#### Elucidating the Potential Impact of circulating Plasmodium falciparum UNIVERSITY **Dihydropteroate Synthase variants on Sulphadoxine-Pyrimethamine Resistance in OF GHANA** Noguchi Memorial Institute **Ghana and Across Africa** for Medical Research Rita Afrivie Boatenq<sup>1</sup>; James Myers-Hansen<sup>1</sup>; Nigel N.O. Dolling<sup>1</sup>; Benedicta Mensah<sup>1</sup>; Brodsky Elia<sup>2</sup>; Mazumder Mohit<sup>2</sup>; Ghansah Anita<sup>1</sup> ogic Bioinformatics 1. Noguchi Memorial Institute for Medical Research, College of Health Sciences, University of Ghana for Infectious Diseases

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### Background

- \* Malaria is still a disease of public health concern globally especially in sub-Saharan Africa
- \* Resistance to Antimalarial therapy is a major concern for malaria control and elimination
- In Ghana, Sulphadoxine-pyrimethamine (SP) is used as Intermittent Preventive Treatment (IPTp) for pregnant women and for children (IPTc)
- SP works by inhibiting Plasmodium dihydropteroate synthase (Pfdhps) and dihydrofolate synthase (Pfdhfr) interfering with folate synthesis
- Initial data analysis of Pfdhps isolates from Ghanaian parasites exhibited a high prevalence of the A581G and A613S in the forest and coastal regions
- This two mutations are associated with resistance to Sulphadoxine and a rare occurrence globally.
- \*Therefore, there is the need to map out Sulphadoxine resistance and assess how the two mutations are spreading across Ghana and other African countries
- Hypothesis: A581G and A613S mutations are spre Ghana and other sub-Saharan African countries one of the driving forces of SP resistance in Africa

# Methodology

#### Data set curation

- \* Here, we curated data from the Plasmodium Community Project (2020) of the MalariaGen repository. The data included 7,113 samples from 29 countries.
- In addition, new data collected in Ghana from two ecologically distinct zones (Begoro and Cape Coast) between 2014-2017 were analysed. The data included 150 and 181 sequences from Begoro and Cape Coast, respectively.





above 20% in Cameroon, Tanzania, Thailand, Cambodia and

Mutation A613S showed variable prevalence across mos Sulphadoxine resistant gene marker A437G was sho

prevalent (almost fixed) in most countries studied

Table 2. Pfdhps haplotypes and their frequencies across Africa

Haplotype		West Africa	East Africa	Central Africa	Southeast Asia	South America
AKAA	Wild-type	4.2-42.6%	0-8.9%	3.3 - 3.8 %	0.2-13.9%	44.8-82.4%
AKAS	Single	0.8 – 2 %	0-0.6 %	0 %	0 %	0 %
GKAA	Single	47.5 - 79.1%	0.4 - 3.3 %	66.7 - 84.7 %	0-38.1%	6.9-11.8%
AKGS	Double	0%	0%	0-0.4 %	0%	0%
GEAA	Double	0.7 - 3.0 %	59.5 - 94.9 %	0-8.5%	17.2-33.0%	0%
GKGA	Double	0%	0%	0-0.3 %	1.3-2.0 %	0-17.2%
GKAS	Double	5.2-11.3%	0-1.6%	0-7.2 %	0-0.1%	5.9 -17.2 %
GKGS	Triple	0-2.1%	0%	0-21.5 %	0%	0%

In total, 17 unique haplotypes on the Pfdhps locus were identified, but 8 were prevalent.

- A single mutation AKAS, double mutant GEAA, triple GKGS, GNGA were most prevalent
- The triple mutant haplotype A437G/A581G/A613S were
- prevalent only in West and Central Africa



### **Results and Discussion**

\* A581G occupies crucial loop structure at the active site in close proximity to A437G and K540E (Fig. 3) \* A613S occurs on alpha helical structure at the entrance of the active site of the protein(Fig. 3) \* Their positions may have a vital effect on sulphadoxine

binding

# Table 2. Effect of mutations on the stability of Pfdhps structure

Mutation	Physiochemical changes	Location on structure	Amino acid changes		
A581G	Hydrophobic to hydrophobic	Buried	Large to small	Destabilizing	0.31
A613S	Hydrophobic to polar	Surface	Small to large	Destabilizing	0.31
A4376	Hydrophobic to hydrophobic	Surface	Large to small	Destabilizing	0.66
K540E	Basic to polar	Surface	Small to large	Destabilizing	0.37
A437G,A613S	1000	Sec. al	1.1.1.1	Stabilizing	-0.06
A437G_A581G	1000	0	Part -	Destabilizing	
A437G, A581G, A613S		243	16 C.	Destabilizing	
A437G, A581G, K540E		and and	Ser.	Destabilizing	2

pilize the active site ould result in conformational changes which



Fig. 5. Drugs and their Binding Energy Profiles in the presence of mutations The presence of mutations caused the loss of molecular interactions surrounding the mutant position

Hydrogen bonds were misplaced indicating major structural

and functional recognition

#### Table 3. Drugs and their Binding Energy Profiles in the presence of mutations

Proteins	Binding energy	Proteins	Binding energ					
NT	-9.3	K540E	-9.3					
4581G	-9.3	A437G,A613S	-9.0					
A613S	-9.4	A437G_A581G	-8.9					
437G	-9.4	A437G, A581G, A613S	-7.8					
		A437G, A581G, K540E	-7.9					

Single mutants were predicted to not significantly decreased binding affinity

BIO

\* Double and triple mutants significantly decrease binding affinity to sulfadoxine



Fig. 6. Molecular Interaction profiles of sulphadoxine in wildtype and mutant proteins

- \*In WT, sulphadoxine made several hydrogen, Pi- and hydrophobic bonds with active site residues crucial for drug recognition and catalysis
- The presence of mutations resulted in change in interactions stabilizing sulphadoxine
- Sulphadoxine formed unfavorable interactions with Tyr663 could resulted in steric clashes

# Conclusion

- \*A581G and A613S mutations are quickly emerging in Ghana and throughout Africa thus increased surveillance is duly recommended
- These mutations occurred on the same haplotype of genetic makers A437G and K540E
- \*A581G and A613S are located on beta and loop region respectively closer to the entrance of the active pocket of the Pfdhps protein
- \*Their presence destabilizes neighboring residues through the loss of molecular interactions
- Initial docking resulted in the loss of molecular interactions between the drug and the binding pocket of the protein
- Unfavorable bonds could suggest steric hindrances between the drug and the residues in the presence of mutations
- Overall, structural analysis suggest potential effect of mutations on drug binding and structure
- \*As future work, running dynamic simulations could additionally highlight some resistance mechanisms